Ammonium to Nitrate Ratio Affects Protocorm Like Bodies PLB Formation In vitro of Hybrid Cymbidium

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Carbon and nitrogen are the two elements that most affect plant organogenesis. In vitro, usually as part of the macronutrients. Some anecdotal evidence from the literature suggests that the ammonium (NH₄⁺) to nitrate (NO₃⁻) ratio may affect orchid organogenesis. In this study, to test this hypothesis, different NH₄⁺: NO₃⁻ ratios were tested on the development of protocorm-like bodies (PLBs) of hybrid Cymbidium Twilight Moon ‘Day Light’, when modified in Teixeira Cymbidium (TC) No. 1 medium without plant growth regulators. More PLBs formed per half-PLB or per PLB tTCL when TC medium was used while TC from which PGRs or NO₃⁻ or NH₄⁺ were removed (two controls) contained significantly less PLBs/explant. The same trend was observed for neo-PLB (i.e., newly formed PLB) fresh weight. A high NO₃⁻ ratio favoured neo-PLB formation, but always less than TC medium, but significantly higher than the two controls. tTCLs performed more poorly (i.e., fewer neo-PLBs/explant and lower fresh weight) than half-PLBs. The NH₄⁺: NO₃⁻ ratio can be manipulated to increase the level of PLB production in the absence of plant growth regulators.

Keywords: Ammonium, Nitrate, PLB, Teixeira Cymbidium (TC) medium.
INTRODUCTION

Nitrogen (N) can be taken up by plants either as inorganic ions (ammonium, NH$_4^+$ or nitrate, NO$_3^-$), or as organic N (Barker, 1975). Among the essential nutrients, N has the greatest impact on plant growth, and tends to be species-specific (Marschner, 1995). Practical applications of this knowledge show that the yield of horticultural crops can be affected by the NH$_4^+$:NO$_3^-$ ratio (Toor et al., 2006). The slow growth of orchids was attributed to a sluggish N metabolism (Poddubnaya-Arnold, 1967). Oyamada (1989) showed that a high NO$_3^-$:NH$_4^+$ ratio resulted in as much as a 10-fold increase in fresh weight when Cymbidium PLBs were placed in liquid shake culture. The medium composition of several plant tissue culture media affected the organogenic outcome in hybrid Cymbidium protocorm-like body (PLB) formation and propagation (Teixeira da Silva et al., 2005). Based on this knowledge, a new medium, Teixeira Cymbidium (TC) No. 1 medium was developed (Teixeira da Silva, 2012), favouring PLB production more than traditionally used Vacin and Went (VW; 1949) medium. In other orchids, high Cl$^-$ and H$^+$ concentrations positively affected seedling regeneration in Phalaenopsis hybrid while increased NH$_4^+$ and NO$_3^-$ concentrations promoted shoot growth (Hinnen et al., 1989). The supply of calcium nitrate to Dendrobium wardianum cultures resulted in direct shoot formation, surpassing the PLB formation stage, as would commonly occur with all the other nitrogen sources (Sharma and Tandon, 1992). Urea, although reported by Mariat (1948) to be an effective N source for Cattleya embryos, is a necrotic agent at higher concentrations (Sharma and Tandon, 1992). Concentrations and NH$_4^+$:NO$_3^-$ ratio in the culture media have been known to affect the germination of orchid seeds and subsequent organogenesis considerably (for example, Shimasaki and Uemoto (1990) for Cymbidium and Zeng et al. (unpublished), for Cypripedium.

In vitro protocols for the induction and development of PLBs of hybrid Cymbidium are well established. Despite this, no study has yet examined the impact of the NH$_4^+$:NO$_3^-$ ratio on organogenesis of Cymbidium. This is also the first study to examine the effect of the NH$_4^+$:NO$_3^-$ ratio on PLB production in any orchid.

MATERIAL AND METHODS

Chemicals and reagents

All chemicals and reagents were of the highest analytical grade available and were purchased from either Sigma-Aldrich (St. Louis, USA), Wako Pure Chemical Industries (Osaka, Japan) or Nacalai Tesque (Kyoto, Japan), unless specified otherwise.

Plant material and culture conditions

The protocol is almost identical to that described by Teixeira da Silva (2012). PLBs of hybrid Cymbidium Twilight Moon ‘Day Light’ (Bio-U, Japan) originally developed spontaneously from shoot-tip culture on Vacin and Went (VW, 1949) agar medium without PGRs, were induced and subcultured every two months on TC medium (Teixeira da Silva, 2012), and was supplemented with two plant growth regulators, or PGRs (0.1 mg/l α-naphthaleneacetic acid and 0.1 mg/l kinetin), 2 g/l tryptone and 20 g/l sucrose, and solidified with 8 g/l Bacto agar (Difco Labs., USA), according to procedures and advice outlined by Teixeira da Silva et al. (2005) and Teixeira da Silva and Tanaka (2006). All media were adjusted to pH 5.3 with 1 N NaOH or HCL prior to autoclaving at 100 kPa for 17 min. Cultures were maintained on 40 ml medium in 100-ml Erlenmeyer flasks, double-capped with aluminium foil, at 25°C, under a 16-h photoperiod with a light intensity of 45 μmol m$^{-2}$ s$^{-1}$ provided by 40W fluorescent lamps (Homo Lux, Matsushita Electric Industrial Co., Japan). Longitudinally bisected PLBs (3-4 mm in diameter), referred throughout as half-PLBs, 10 per flask, were used as the explants for PLB induction and proliferation and as one of the two explants for all experiments. The second explant used was PLB transverse thin cell layers or tTCLs, prepared as explained in detail by Teixeira da Silva (2013). Culture conditions and media followed
the recommendations previously established for medium formulation (Teixeira da Silva et al., 2005), biotic (Teixeira da Silva et al., 2006b) and abiotic factors (Teixeira da Silva et al., 2006a) for PLB induction, formation and proliferation.

**Effect of NH$_4^{+}$:NO$_3^{-}$ ratio on PLB formation**

Half-PLBs and PLB tTCLs were cultured on TC medium containing one of the following NH$_4^{+}$:NO$_3^{-}$ ratios: 0:10, 2.5:7.5, 1:1, 7.5:2.5, 10:0, all at the same N molarity as TC. Two controls were used: C-PGR, which contained no PGRs but contained TC N levels and C-N, which contained TC PGRs but no NO$_3^{-}$ or NH$_4^{+}$. The number of PLBs formed per PLB segment or PLB tTCL was measured. All measurements were made after 45 days in culture.

**Statistical analyses**

Experiments were organized according to a randomized complete block design (RCBD) with three blocks of 10 replicates per treatment. All experiments were repeated in triplicate (n = 90, total sample size per treatment). Data was subjected to analysis of variance (ANOVA) with mean separation by Duncan’s New Multiple Range Test (DNMRT) using SAS® version 6.12 (SAS Institute, Cary, NC, USA). Significant differences between means were assumed at p ≤ 0.05.

**RESULTS AND DISCUSSION**

TC medium resulted in the highest number of PLBs per half-PLB or per PLB tTCL while TC from which PGRs or NO$_3^{-}$ or NH$_4^{+}$ were removed contained significantly less PLBs/explant (Table 1). The same trend was observed for neo-PLB fresh weight. A neo-PLB is a newly formed PLB. A high ratio of NO$_3^{-}$ favoured neo-PLB formation, but never higher than TC medium, but significantly higher than both control treatments (Table 1). tTCLs performed more poorly (i.e., fewer neo-PLBs/explant and lower fresh weight) than half-PLBs, but this is understandable considering that tTCLs have a much lower surface area than half-PLBs (Teixeira da Silva and Dobránszki, 2013). Despite lower apparent, face-value productivity levels of tTCLs, it is likely that true productivity could exceed that of half-PLBs once a suitable conversion factor, the plant Growth Correction Factor, is applied (Teixeira da Silva and Dobránszki, 2011).

Kubota et al. (2000) noted that NH$_4^{+}$ antagonistically inhibited the absorption of P, K, Ca and Mg, but particularly divalent cations Ca$^{2+}$ and Mg$^{2+}$; moreover, when NO$_3^{-}$ was used as the only N source, or when the ratio of NO$_3^{-}$ was high, then *Phalaenopsis* plant growth was favoured. In their study, Kubota et al. (2000) also noted that leaf number decreased at a 8:2 and a 10:0 ratio of NH$_4^{+}$:NO$_3^{-}$, leaf area increased when the ratio was 0:10 and 2:8, and root number and length increased linearly as the NH$_4^{+}$ concentration increased, but leaf, root and whole plant dry weight decreased as the NH$_4^{+}$ concentration increased. Hinnen et al. (1989) had recommended a 6:4 ratio of NH$_4^{+}$:NO$_3^{-}$ for optimal growth of *Phalaenopsis* shoots, which contradicted the findings of Kubota et al. (2000). In the commercial production of orchids, Hyponex, a commercial liquid fertilizer, or in powder form, is used, with an N-P-K of 6.5-2.6-15.8, and this usually contains 14 mM of N, consisting of a 1.5:8.5 NH$_4^{+}$:NO$_3^{-}$ ratio (Ichihashi, 1997), favouring the need for NO$_3^{-}$, as also presumed by Kubota et al. (2000). Ichihashi (1979) indicated that a 4:6 NH$_4^{+}$:NO$_3^{-}$ ratio improved the *In vitro* growth of *Bletilla striata* seedling growth, but since molarities differed, those results were in fact flawed and/or inconclusive. Studying two epiphytic orchids, *Dendrobium nobile* and *Cattleya*, Uesato (1973, 1974) showed that a 8:2 or 7:3 ratio of NH$_4^{+}$:NO$_3^{-}$, respectively promoted *In vitro* shoot growth, but that shoot growth was depressed when the NH$_4^{+}$ ratio increased further. Shimasaki and Uemoto (1990) noted that high levels of NH$_4^{+}$NO$_3^{-}$ (ammonium nitrate) inhibited the growth of *Cymbidium kanran* (a terrestrial orchid) shoots *In vitro*, but that moderate levels of NH$_4^{+}$NO$_3^{-}$ together with moderate levels of potassium nitrate (KNO$_3$) increased shoot and root growth. Using nutrient solutions, Hew et al. (1992) showed that the growth of two terrestrial or-
chids, *Cymbidium* and *Bromheadia*, was not perturbed if only NH$_4^+$ was used. These studies that exist on the manipulation of the NH$_4^+$:NO$_3^−$ ratio in orchids indicates that the optimal NH$_4^+$:NO$_3^−$ ratio depends on the species and even on the genotype (cultivar).

In other crops *In vitro*, a high NH$_4^+$:NO$_3^−$ ratio improved garlic (*Allium sativum*) micropropagation (Luciani *et al*., 2001). By changing the NH$_4^+$:NO$_3^−$ ratio from 39.5 mM + 20.5 mM to 15.8 mM + 20.5 mM, Bensaddek *et al.* (2001) could improve the growth, alkaloid production and scopalamine/hyoscyamine ratio of two clones of belladonna (*Atropa belladonna*) hairy roots. By exposing plants to a high level of ammonium nitrate, Greer *et al.* (2009) were able to improve the frequency of wheat (*Triticum aestivum*) transformation. Garnica *et al.* (2010) showed that NO$_3^−$ reduces the negative effects caused by NH$_4^+$ and promotes plant growth by enhancing the levels of cytokinins and indole-3-acetic acid, and lowering the levels of abscisic acid. There are many other studies that have examined the effect of the NH$_4^+$:NO$_3^−$ ratio in greenhouse or field production, but those studies lie beyond the scope of this study.

PLBs and somatic embryos are synonymous in orchids (Teixeira da Silva and Tanaka, 2006), and thus this protocol represents another simple way of producing PLBs, which are essential to orchid biotechnology (Hossain *et al*., 2013).

**ACKNOWLEDGEMENTS AND CONFLICT OF INTEREST**

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In vitro-A review. In review.
## Table 1. Effect of NH₄⁺:NO₃⁻ ratio on PLB formation from half-PLB culture of hybrid *Cymbidium Twilight Moon* ‘Day Light’.

<table>
<thead>
<tr>
<th>Medium composition</th>
<th>NH₄⁺:NO₃⁻ ratio (all identical molarity)</th>
<th>Percentage of explants forming neo-PLBs (%)</th>
<th>Number of PLBs per explant</th>
<th>Fresh weight (mg) of PLB explant + neo-PLBs</th>
</tr>
</thead>
<tbody>
<tr>
<td>TC medium*</td>
<td>100 a</td>
<td>8.3 a</td>
<td>526 a</td>
<td></td>
</tr>
<tr>
<td>TC medium**</td>
<td>100 a</td>
<td>1.6 de</td>
<td>103 e</td>
<td></td>
</tr>
<tr>
<td>C-PGR (control 1)*</td>
<td>47 c</td>
<td>1.4 de</td>
<td>108 e</td>
<td></td>
</tr>
<tr>
<td>C-N (control 2)*</td>
<td>23 a</td>
<td>0.8 e</td>
<td>86 f</td>
<td></td>
</tr>
<tr>
<td>Half-PLBs</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>0:10</td>
<td>96 a</td>
<td>7.1 b</td>
<td>418 b</td>
<td></td>
</tr>
<tr>
<td>2.5:7.5</td>
<td>91 ab</td>
<td>6.3 c</td>
<td>381 c</td>
<td></td>
</tr>
<tr>
<td>1:1</td>
<td>76 b</td>
<td>2.6 d</td>
<td>196 d</td>
<td></td>
</tr>
<tr>
<td>7.5:2.5</td>
<td>27 d</td>
<td>1.1 e</td>
<td>83 f</td>
<td></td>
</tr>
<tr>
<td>10:0</td>
<td>0 f</td>
<td>0 f</td>
<td>0 h</td>
<td></td>
</tr>
<tr>
<td>PLB tTCLs</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>0:10</td>
<td>88 ab</td>
<td>1.4 de</td>
<td>74 fg</td>
<td></td>
</tr>
<tr>
<td>2.5:7.5</td>
<td>61 bc</td>
<td>1.1 e</td>
<td>56 g</td>
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<tr>
<td>1:1</td>
<td>45 c</td>
<td>0.6 ef</td>
<td>48 g</td>
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<tr>
<td>7.5:2.5</td>
<td>13 e</td>
<td>0.1 f</td>
<td>26 gh</td>
<td></td>
</tr>
<tr>
<td>10:0</td>
<td>0 f</td>
<td>0 f</td>
<td>0 h</td>
<td></td>
</tr>
</tbody>
</table>

Mean values followed by the same letter in the same column are not significantly different based on DMRT (P = 0.05). n = 90 (9 Petri dishes x 10 for each treatment). * = half-PLBs; ** = tTCLs. C-PGR: contains no PGRs but contains TC N levels; C-N, contains TC PGRs but no NO₃⁻ or NH₄⁺. General abbreviations: PGR, plant growth regulator; PLB, protocorm-like body; TC, Teixeira *Cymbidium* No. 1 medium (Teixeira da Silva 2012), includes 0.1 mg/l α-naphthaleneacetic acid and 0.1 mg/l kinetin, 2 g/l tryptone and 20 g/l sucrose (see reference for modified micro- and macro-nutrients); tTCL, transverse thin cell layer.